Crystallins as Important Pathogenic Targets for Accumulation of Structural Damages Resulting in Protein Aggregation and Cataract Development: Introduction to This Special Issue of *Biochemistry (Moscow)*

Reza Yousefi^{1,2}

¹Institute of Biochemistry and Biophysics, University of Tehran, Tehran, Iran
²Protein Chemistry Laboratory, College of Sciences, Shiraz University, Shiraz, Iran
e-mail: yousefi.reza@ut.ac.ir, ryousefi@shirazu.ac.ir

Received January 29, 2022 Revised January 29, 2022 Accepted January 29, 2022

Abstract—This issue of *Biochemistry (Moscow)* is dedicated to the role of protein misfolding and aggregation in cataract development. In fact, many genetic mutations or chemical and physical deleterious factors can initiate alterations in the macrostructural order and proper folding of eye lens proteins, which in some cases result in the formation of large light-scattering aggregates, affecting the quality of vision and making lens more prone to cataract development. Diabetes mellitus, which is associated with oxidative stress and mass production of highly reactive compounds, can accelerate unfolding and aggregation of eye lens proteins. This journal issue contains reviews and research articles that describe the destructive effects of mutations and highly reactive metabolites on the structure and function of lens crystallin proteins, as well important molecules in the lens's natural defense system involved in protection against deleterious effects of the physical and chemical factors.

DOI: 10.1134/S0006297922020018

Keywords: lens proteins, mutations, diabetes mellitus, reactive substances, lens natural defense system

INTRODUCTION

This issue, which is dedicated to protein misfolding and aggregation in cataract disorders, was prepared at the suggestion and with participation of Boris Ivanovich Kurganov, Chief Researcher of the Bach Institute of Biochemistry and Honored Scientist of the Russian Federation, who passed away on October 1, 2021. His unremitting research efforts in the field of enzymology, mechanisms of protein aggregation, and role of molecular and chemical chaperones in prevention of protein aggregation, which spanned more than six decades, have left an important and valuable scientific heritage. Boris Kurganov had also played a prominent role in studying the mechanism of aggregation of eye lens crystallins, proteins that have an essential role in the transparency and refractive index of the lenticular tissues [1].

CRYSTALLIN PROTEINS AS IMPORTANT PATHOGENIC TARGETS FOR ACCUMULATION OF VARIOUS STRUCTURAL DAMAGES

These water-soluble structural proteins found in the vertebrate eye lenses are classified into three main types $-\alpha$ -, β -, and γ -crystallins - that elute from a gel filtration column in the same order due to the differences in their size and degree of oligomerization [2]. α -crystallins (αA and αB) and β -crystallins ($\beta A1$, $\beta A2$, $\beta A3$, βA4, βB1, βB2, βB3; A, acidic; B, basic) form larger and smaller oligomers, respectively, while γ -crystallins (γA , γB , γC , γD , γE , γF , γN , and γS) are essentially monomers [3]. These highly stable β -rich proteins form very regular and interactive macrostructures through subtle interactions with each other which are important for lens transparency [1, 4]. In addition to disrupting such important fine protein-protein interactions, genetic mutations and accumulation of physical and chemical damages over the lifespan result in structural changes and

^{*} To whom correspondence should be addressed.

88 YOUSEFI

exposure of protein hydrophobic regions. Eventually, these adverse molecular events lead to the unfolding and aggregation of lens crystallins, inducing cataract development. Cataract is the most common cause of blindness worldwide, affecting tens of millions of people [5-9]. Beside lenticular tissues, crystallin proteins (e.g., αBcrystallin) are found in other tissues, such as retina, heart, skeletal muscle, skin, brain, etc. [10, 11]. Therefore, structural and functional damages to these proteins are also associated with other disorders, including myopathy, neurological problems, cardiac diseases, muscular disorders, and invasive breast cancer tumors [12-15]. Along with the structural role, crystallins have several metabolic and regulatory functions, both inside and outside of lenticular tissues [16]. Although enzymatic activity has been reported for some type of crystallins [17], their most important biological function can be attributed to the chaperone activity of α -crystallin, which plays a vital role in preventing and delaying cataract diseases and increases cell resistance to various types of chemical and physical stress [18, 19]. This chaperone, which belongs to the heat shock protein (hsp) family, also plays a prominent role in the inhibition of apoptosis and cytoskeletal remodeling [20, 21]. Crystallins are highly clear and refractive proteins with abnormal hydration properties. They resist unfolding and aggregation for decades during human life [22]. However, mutations and accumulation of physical and chemical damages eventually cause unfolding and aggregation of these proteins [23]. Also, some diseases (e.g., diabetes) characterized by the elevated levels of oxidative compounds and reactive metabolites (sugars and sugar derivatives) in eye lenses, increase the extent of crystallin structural damage and thus accelerate the process of cataract formation [24, 25]. Mutations in the genes of two α-crystallin subunits are also associated with the dominant and recessive forms of cataract, as well as with a wide range of neurological, cardiovascular, and muscular disorders [26-28]. Mutations that cause severe damage to these proteins usually result in congenital cataracts, while milder mutation increase lens susceptibility to the environmental damage and are associated with the age-related cataract development [29]. Also, with age, gradual accumulation of covalent damages due to various factors, such as ultraviolet radiation [30], oxidation [31], deamidation [32], and proteolysis [33], results in the formation of protein aggregates that scatter the incident light in the lens. Numerous studies have also demonstrated that the abnormal levels of essential elements (calcium, copper, and zinc) and heavy metals (divalent lead, cadmium, and mercury) are the important sources of destructive damages to crystallin proteins and, under some circumstances, can be considered as potential causative factors in cataract development [34-37]. For example, diseases that increase the level of essential metals in eye lenses (e.g., diabetes) also cause structural damage to lens proteins and induce their aggregation, which

further facilitates the development of cataract complications [38, 39]. Although, the only currently available treatment is surgical removal of cataract lenses, scientific and medical community has long sought for the non-destructive treatments as well. In this regard, therapies based on the use of natural products [40], modulators of oxidation processes [41], protein aggregation inhibitors (e.g., chemical chaperones) [42], homoeopathic remedies [43], and lens regeneration using endogenous stem cells [44] have been proposed.

Below, I will briefly introduce the articles of this special issue [51, 53, 57, 58, 60]. Over the years, Professor Boris Kurganov's research team has developed methods for evaluating the effect of molecular and chemical chaperones, as well as the effects of their combined application on the kinetics of protein aggregation [45-50]. In continuation of these studies, Chebotareva et al. [51] investigated the effect of trehalose as a chemical chaperone, on the quaternary structure and chaperone activity of αB-crystallin. Mature fiber cells contain extremely high concentrations of crystallin proteins that make up approximately 90% of the dry weight of human lens [52]. While lens proteins are continuously exposed to physical and chemical damage, lens cells have developed protective systems to counteract the harmful effects of environmental factors on lens proteins. However, in the case of diseases, such as diabetes or aging, the dominance of the damaging factors over two important levels of natural protections (chaperone and antioxidant defense systems) can have serious destructive effects on the structure and function of lens proteins. Various approaches for preventing lens opacity, in particular, combined use of antioxidants and chemical molecules, have been reviewed by Muranov and Ostrovsky [53]. Diabetes mellitus is one of the causes of rapid cataract development. Beside oxidative stress, this metabolic disorder is characterized by the elevated concentrations of reactive metabolites, such as glucose, fructose, phosphorylated sugars (glycolytic intermediates), methylglyoxal, peroxynitrite, and sorbitol, in the lenticular tissues [54]. The oxidative stress typical for diabetes promotes the reaction between sugars or sugar derivatives and eye lens proteins [55]. The osmotic stress created by the increased sorbitol accumulation in eye lenses in hyperglycemia is one of the mechanisms of diabetic cataract [56]. Therefore, the effect of different concentrations of sorbitol on the structure and chaperone-like activity of rat α -crystallin has been studied by Reddy et al. [57]. The impact of peroxynitrite (an important source of oxidative stress in diabetes), methylglyoxal (diabetes-associated reactive carbonyl compound), and their simultaneous action on the structure and function of human recombinant αA -crystallin (αA -Cry) and the protective role of ascorbic acid and glutathione (main components of lens antioxidant defense system) have been investigated by Yousefi et al. [58]. Many mutations that found in the crystallin protein

genes are associated with diseases such as cataracts and myopathy [59]. Finally, the role of genetic mutations of α -crystallins on their structural unfolding and aggregation is discussed by Rao et al. [60].

This issue was initiated by and until recently created with a significant contribution from Professor Kurganov, and now it had to be finished in his absence. This is why we dedicate it to the memory of Professor Boris Ivanovich Kurganov, whose relentless efforts over the past decades and his valuable and influential scientific legacy have paved the way for other researchers to the unknown frontiers of knowledge.

Funding. This work was supported by the Iran National Science Foundation, INSF (Grant No. 99014455).

Ethics declarations. The author declares no conflicts of interest. This article does not contain description of studies with the involvement of humans or animal subjects performed by the author.

REFERENCES

- Delaye, M. (1983) Short-range order of crystallin proteins accounts for eye lens transparency, *Nature*, 302, 415-417.
- Bloemendal, H. (1977) The vertebrate eye lens, *Science*, 197, 127-138.
- 3. Lampi, K. J., Ma, Z., Shih, M., Shearer, T. R., Smith, J. B., et al. (1997) Sequence analysis of betaA3, betaB3, and betaA4 crystallins completes the identification of the major proteins in young human lens, *J. Biol. Chem.*, 272, 2268-2275.
- 4. Benedek, G. B. (1971) Theory of transparency of the eye, *Appl. Opt.*, **10**, 459-473.
- 5. Bera, S., and Abraham, E. C. (2002) The αA-crystallin R116C mutant has a higher affinity for forming heteroaggregates with αB-crystallin, *Biochemistry*, **41**, 297-305.
- Basha, E., O'Neill, H., and Vierling, E. (2012) Small heat shock proteins and α-crystallins: Dynamic proteins with flexible functions, *Trends Biochem. Sci.*, 37, 106-117.
- Bron, A. J., Vrensen, G., Koretz, J., Maraini, G., and Harding, J. (2000) The ageing lens, *Opthalmologica*, 214, 86-104
- 8. Takemoto, L. J., and Ponce, A. A. (2006) Decreased association of aged alpha crystallins with gamma crystallins, *Exp. Eye Res.*, **83**, 793-797.
- Treweek, T. M., Rekas, A., Lindner, R. A., Walker, M. J., Aquilina, J. A., et al. (2005) R120G αB-crystallin promotes the unfolding of reduced α-lactalbumin and is inherently unstable, *FEBS J.*, 272, 711-724.
- Dubin, R. A., Wawrousek, E. F., and Piatigorsky, J. (1989)
 Expression of the Murine αB-crystallin gene is not restricted to the lens, *Mol. Cell Biol.*, 9, 1083-1091.
- 11. Sax, C. M., and Piatigorsky, J. (1994) Expression of the alpha-crystallin/small heat-shock protein/molecular chaperone genes in the lens and other tissues, *Adv. Enzymol. Relat. Areas Mol. Biol.*, **69**, 155-201.
- 12. Koletsa, T., Stavridi, F., Bobos, M., Kostopoulos, I., Kotoula, V., et al. (2014) alphaB-crystallin is a marker of

- aggressive breast cancer behavior but does not independently predict for patient outcome: A combined analysis of two randomized studies, *BMC Clin. Pathol.*, **14**, 1-13.
- 13. Rajasekaran, N. S., Connell, P., Christians, E. S., Yan, L., Taylor, R. P., et al. (2007) Human alphaB-crystallin mutation causes oxido-reductive stress and protein aggregation cardiomyopathy in mice, *Cell*, **130**, 427-439.
- Simon, S., Fontaine, J., Martin, J. L., Sun, X., Hoppe, A. D., et al. (2007) Myopathy-associated alphaB-crystallin Mutants abnormal phosphorylation, intracellular location, and interactions with other small heat shock proteins, *J. Biol. Chem.*, 282, 34276-34287.
- Avliyakulov, N. K., Rajavel, K. S., Minh, K., Haykinson, M. J., and Pope, W. B. (2014) C-terminally truncated form of alphaB-crystallin is associated with IDH1 R132H mutation in anaplastic astrocytoma, *J. Neurooncol.*, 117, 53-65.
- Boelens, W. C. (2014) Cell biological roles of αB-crystallin, *Prog. Biophys. Mol. Biol.*, 115, 3-10.
- Wistow, G., and Kim, H. (1991) Lens protein expression in mammals: Taxon-specificity and the recruitment of crystallins, J. Mol. Evol., 32, 262-269.
- 18. Horwitz, J. (1992) a-Crystallin can function as a molecular chaperone, *Proc. Natl. Acad. Sci. USA*, **89**, 10449-10453.
- Andley, U. P. (2007) Crystallins in the eye: Function and pathology, *Prog. Retin. Eye Res.*, 26, 78-98.
- 20. Van Montfort, R., Slingsby, C., and Vierlingt, E. (2001) structure and function of the small heat shock protein/α-crystallin family of molecular chaperones, *Adv. Protein Chem.*, **59**, 105-156.
- Pasupuleti, N., Matsuyama, S., Voss, O., Doseff, A. I., Song, K., et al. (2010) The anti-apoptotic function of human αA-crystallin is directly related to its chaperone activity, *Cell Death Dis.*, 1, e31-e31.
- Roskamp, K. W., Paulson, C. N., Brubaker, W. D., and Martin, R. W. (2020) Function and aggregation in structural eye lens crystallins, *Acc. Chem. Res.*, 53, 863-874.
- 23. Clark, A. R., Lubsen, N. H., and Slingsby, C. (2012) sHSP in the eye lens: Crystallin mutations, cataract and proteostasis, *Int. J. Biochem. Cell Biol.*, **44**, 1687-1697.
- 24. Yousefi, R., Javadi, S., Amirghofran, S., Oryan, A., and Moosavi-movahedi, A. A. (2016) Assessment of structure, stability and aggregation of soluble lens proteins and alphacrystallin upon non-enzymatic glycation: The pathomechanisms underlying cataract development in diabetic patients, *Int. J. Biol. Macromol.*, 82, 328-338.
- Zafaranchi, S., Khoshaman, K., Masoudi, R., Hemmateenejad, B., and Yousefi, R. (2017) The structural alteration and aggregation propensity of glycated lens crystallins in the presence of calcium: Importance of lens calcium homeostasis in development of diabetic cataracts, *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.*, 170, 174-183.
- Dimauro, I., Antonioni, A., Mercatelli, N., and Caporossi, D. (2018) The role of αB-crystallin in skeletal and cardiac muscle tissues, *Cell Stress Chaperones*, 23, 491-505
- Shiels, A., Hejtmancik, J. F., Sciences, V., and Branch, V. F. (2017) Mutations and mechanisms in congenital and age-related cataracts, *Exp. Eye Res.*, 156, 95-102.
- Phadte, A. S., Sluzala, Z. B., and Fort, P. E. (2021)
 Therapeutic potential of α-crystallins in retinal neurodegenerative diseases, *Antioxidants*, 10, 1-13.

90 YOUSEFI

 Pescosolido, N., Barbato, A., Giannotti, R., Komaiha, C., and Lenarduzzi, F. (2016) Age-related changes in the kinetics of human lenses: Prevention of the cataract, *Int. J. Ophthalmol.*, 9, 1506.

- Varma, S. D., Kovtun, S., and Hegde, K. R. (2011) Role of UV irradiation and oxidative stress in cataract formation. Medical prevention by nutritional antioxidants and metabolic agonists, *Eye Contact Lens*, 37, 233-245.
- 31. Linetsky, M., Shipova, E., Cheng, R., and Ortwerth, B. J. (2008) Glycation by ascorbic acid oxidation products leads to the aggregation of lens proteins, *Biochim. Biophys. Acta*, **1782**, 22-34.
- Pande, A., Mokhor, N., Pande, J., and States, U. (2018)
 Deamidation of human γS-crystallin increases attractive protein interactions: Implications for cataract, *Biochemistry*, 54, 4890-4899.
- 33. Gong, X., Li, E., Klier, G., Huang, Q., Wu, Y., et al. (1997) Disruption of alpha3 connexin gene leads to proteolysis and cataractogenesis in mice, *Cell*, **91**, 833-843.
- 34. Kashani, M. R., Yousefi, R., Akbarian, M., Alavianmehr, M. M., and Ghasemi, Y. (2016) Structure, chaperone activity, and aggregation of wild type and R12C mutant αB crystallins in the presence of thermal stress and calcium ion implications for role of calcium in cataract pathogenesis, *Biochemistry*, 81, 122-134.
- Ghahramani, M., Yousefi, R., Khoshaman, K., Sasan, S., and Kurganov, B. I. (2016) Evaluation of structure, chaperone-like activity and protective ability of peroxynitrite modified human alpha-Crystallin subunits against coppermediated ascorbic acid oxidation, *Int. J. Biol. Macromol.*, 87, 208-221.
- Calva, J. A. D., Vázquez, M. L. P., and King, J. A., and Quintanar, L. (2018) Mercury-induced aggregation of human lens γ-crystallins reveals a potential role in cataract disease, J. Biol. Inorg. Chem., 23, 1105-1118.
- Kempka, K., Kaminski, P., Malukiewicz, G., Bogdzinska, M., and Florczak, S. (2018) Initial proantioxidant reactions in the patients suffering from cataract in the interactions with cadmium and lead, World Sci., 108, 195-206
- 38. Kyselova, Z., Stefek, M., and Bauer, V. (2004) Pharmacological prevention of diabetic cataract, *J. Diabetes Complicat.*, **18**, 129-140.
- Moreau, K. L., and King, J. A. (2012) Protein misfolding and aggregation in cataract disease and prospects for prevention, *Trends Mol. Med.*, 18, 273-282.
- 40. Kato, K., Ito, H., Kamei, K., and Iwamoto, I. (1998) Stimulation of the stress-induced expression of stress proteins by curcumin in cultured cells and in rat tissues *in vivo*, *Cell Stress Chaperones*, **3**, 152.
- 41. Khoshaman, K., Yousefi, R., and Moosavi-movahedi, A. A. (2017) Protective role of antioxidant compounds against peroxynitrite-mediated modification of R54C mutant a A-crystallin, *Arch. Biochem. Biophys.*, **629**, 43-53.
- Jara, O., Minogue, P. J., Berthoud, V. M., and Beyer, E. C. (2018) Chemical chaperone treatment improves levels and distributions of connexins in Cx50D47A mouse lenses, *Exp. Eye Res.*, 175, 192-198.
- 43. Lian, R. R., and Afshari, N. A. (2020) The quest for home-opathic and nonsurgical cataract treatment, *Curr. Opin. Ophthalmol.*, **31**, 61-66.

- 44. Liu, Z., Wang, R., Lin, H., and Liu, Y. (2020) Lens regeneration in humans: using regenerative potential for tissue repairing, *Ann. Transl. Med.*, **8**, 1-17.
- 45. Mikhaylova, V., Eronina, T., and Kurganov, B. (2021) the effect of chemical chaperones on test systems with different kinetic regime of aggregation, *FEBS Open Bio*, **11**, 170-170.
- Ghahramani, M., Yousefi, R., Krivandin, A., Muranov, K., Kurganov, B., et al. (2020) Kinetic data analysis of chaperone-like activity of Wt, R69C and D109H αB-crystallins, Data in Brief, 28, 104922.
- 47. Kurganov, B. I. (2017) Quantification of anti-aggregation activity of chaperones, *Int. J. Biol. Macromol.*, **100**, 104-117.
- 48. Kurganov, B. I. (2015) Selection of test systems for estimation of anti-aggregation activity of molecular chaperones, *Biochem. Anal. Biochem.*, **4**, 1.
- 49. Kurganov, B. I. (2014) Estimation of chaperone-like activity using test systems based on protein amyloid aggregation, *Biochem. Anal. Biochem.*, **4**, 2161-1009.
- Borzova, V. A., Markossian, K. A., Kara, D. A., Chebotareva, N. A., Makeeva, V. F., et al. (2013) Quantification of anti-aggregation activity of chaperones: A test-system based on dithiothreitol-induced aggregation of bovine serum albumin, *PLoS One*, 8, e74367.
- Chebotareva, N. A., Eronina, T. B., Mikhaylova, V., Roman, S. G., Tugaeva, K. V., et al. (2022) Effect of trehalose on oligomeric state and anti-aggregation activity of αB-Crystallin, *Biochemistry (Moscow)*, 87, 121-130.
- Sharma, K. K., and Santhoshkumar, P. (2009) Lens aging: Effects of crystallins, *Biochim. Biophys. Acta*, 1790, 1095-1108.
- 53. Muranov, K. O., and Ostrovsky, M. O. (2022) Lens biochemistry in the norm and in cataractogenesis, *Biochemistry (Moscow)*, **87**, 106-120.
- 54. Moghadam, S. S., Oryan, A., Kurganov, B. I., Tamaddon, A. M., Alavianehr, M. M., et al. (2017) The structural damages of lens crystallins with peroxynitrite and methylglyoxal, two causetive players in diabetic complications and preventive role of lens antioxidant components, *Int. J. Biol. Macromol.*, 103, 74-88.
- 55. Stitt, A. (2005) The maillard reaction in eye disease, *Ann. N. Y. Acad. Sci.*, 1043, 582-597.
- Patel, D. K., Prasad, S. K., Kumar, R., and Hemalatha, S. (2011) Cataract: A major secondary complication of diabetes, its epidemiology and an overview on major medicinal plants screened for anticataract activity, *Asian Pac. J. Trop. Dis.*, 1, 323-329.
- 57. Kumar, C. U., Suryavanshi, U., Sontake, V., Reddy, P. Y., Sankhala, R. S., et al. (2022) Effects of sorbitol on alphacrystallin structure and function, *Biochemistry (Moscow)*, 87, 131-140.
- 58. Moghadam, S. S., Ghahramani, M., Khoshaman, K., Oryan, A., Moosavi-Movahedi, A. A., et al. (2022) Relationship between the structure and chaperone activity of human αA-Crystallin after its modification with diabetes-associated oxidative agents and protective role of antioxidant compounds, *Biochemistry (Moscow)*, 87, 91-105.
- 59. Graw, J. (2009) Genetics of crystallins: Cataract and beyond, *Exp. Eye Res.*, **88**, 173-189.
- 60. Budnar, B., Tangirala, R., Bakthisaran, R., and Rao, C. M. (2022) Protein aggregation and cataract: Role of age-related modifications and mutations in α-crystallins, *Biochemistry (Moscow)*, in press.