

# Bioactive Amide of Prostaglandin E<sub>1</sub> and Ethanolamine Plasmalogen Analog of Platelet-Activating Factor Inhibits Several Pathways of Human Platelet Aggregation

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**Abstract**—The influence of an amide of prostaglandin E<sub>1</sub> and ethanolamine plasmalogen platelet-activating factor analog 1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phospho-(N-11 $\alpha$ ,15 $\alpha$ -dioxy-9-keto-13-prostenoyl)ethanolamine (PGE<sub>1</sub>-PPAF) on platelet-activating factor (PAF)-, ADP-, and thrombin-induced human platelet aggregation has been studied. It was found that PGE<sub>1</sub>-PPAF inhibits the PAF-, ADP-, and thrombin-induced platelet aggregation in platelet-rich plasma. 1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phosphoethanolamine inhibited PAF-induced aggregation up to 50% but had no influence on platelet aggregation induced by ADP or thrombin. The ethanolamine plasmalogen analog of PAF 1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phospho-(N-palmitoyl)ethanolamine, having a palmitoyl residue instead of PGE<sub>1</sub>, did not inhibit platelet aggregation induced by PAF, ADP, or thrombin. We propose that inhibition of human platelet aggregation by PGE<sub>1</sub>-PPAF is mediated by its action on platelet PAF-receptors and the adenylate cyclase system.

**Key words:** platelet-activating factor, amide of prostaglandin E<sub>1</sub> and a PAF analog, prostaglandin E<sub>1</sub>, platelets

Ethanolamides of fatty acids are a new group of lipid bioregulators that are considered as endogenous ligands of cannabinoid receptors of the central nervous system and peripheral tissues [1]. One of the well-studied fatty acid ethanolamides is anandamide—the ethanolamide of arachidonic acid [1]. Cell cyclooxygenase-2 can synthesize the ethanolamide of prostaglandin E<sub>2</sub> from anandamide, and 12- or 15-lipoxygenase can synthesize the 12- or 15-hydroxy derivatives of anandamide; these have a different spectrum of biological activity than anandamide [2].

The hydrolysis of N-acylated phosphatidylethanolamines is one of the pathways of biosynthesis of fatty acid ethanolamides [1]. N-Acylphosphatidylethanolamines have been identified in a number of cells and tissues [3] and obtained by chemical synthesis [4]. Previously, the phospholipid derivatives of prostaglandins having prostaglandins joined by an amide bond with the ethanolamine residue of phos-

phatidylethanolamine were obtained [5]. These phospholipid derivatives of prostaglandins revealed a broad spectrum of biological activity, more stable against inactivation in plasma and tissues than prostaglandins, and are of interest as precursors of bioactive prostaglandin ethanolamides.

In this work, we studied the influence of prostaglandin E<sub>1</sub> and an ethanolamine plasmalogen analog of PAF on human platelet aggregation induced by PAF, ADP, or thrombin. We chose this compound for our study for the following reasons. Under physiological conditions in the bloodstream, platelet activation is not induced by a single agent, but it is the result of the action of a combination of different proaggregatory agents. A great number of proaggregatory agents operate by three general mechanisms involving ADP, thromboxane A<sub>2</sub>, and PAF [6]. The selective or simultaneous action of these three general aggregation mechanisms give rise to the possibility of direct regulation of the functional activity of platelets. It was shown previously that a plasmalogenic analog of PAF (1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phosphocholine) binds with PAF-receptors, induces platelet desensitization to PAF, inhibits PAF-induced platelet desensitization to PAF, and inhibits PAF-induced platelet aggregation, but it has no effect

**Abbreviations:** PAF) platelet-activating factor (1-O-alkyl-2-acetyl-sn-glycero-3-phosphocholine); PGE<sub>1</sub>) prostaglandin E<sub>1</sub>; PGE<sub>1</sub>-PPAF) 1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phospho-(N-11 $\alpha$ ,15 $\alpha$ -dioxy-9-keto-13-prostenoyl)ethanolamine.

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on ADP- or thrombin-induced aggregation [7]. In contrast to PAF, the plasmalogenic analog of PAF increases [ $^3\text{H}$ ]PGE<sub>1</sub> binding with human platelets [7].

It is known that PGE<sub>1</sub> and prostacyclin activate platelet adenylate cyclase by a receptor-dependent route that increases platelet cAMP level and inhibits platelet aggregation, and this effect can be increased by phosphodiesterase inhibitors [8]. Since the plasmalogenic analog of PAF (1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phosphocholine) blocks PAF-receptors and the PAF-dependent aggregation pathway, and PGE<sub>1</sub> inhibits platelet aggregation due to adenylate cyclase activation, we combine these two functions in the PGE<sub>1</sub>-PPAF compound that is aimed at simultaneous action on two platelet receptor systems and inhibition of platelet aggregation induced by different proaggregatory agents.

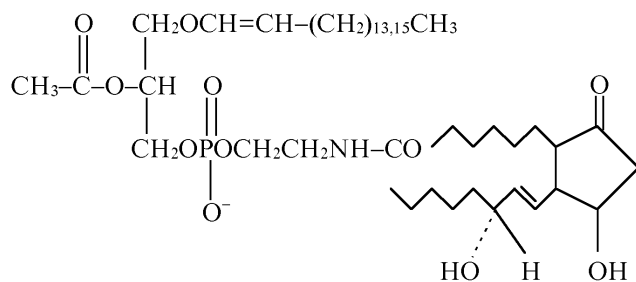
## MATERIALS AND METHODS

PGE<sub>1</sub> and thrombin from Sigma (USA) and ADP from Reanal (Hungary) were used in this study. PAF was obtained from beef heart choline plasmalogens by a classic method [9].

1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phosphoethanolamine (compound 1) was obtained by phospholipase D-catalyzed transesterification of 1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phosphocholine in the presence of ethanolamine as described previously [4].

1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phospho-(N-palmitoyl)ethanolamine (compound 2) was obtained by treatment of compound 1 with palmitic anhydride in benzene and purified by column chromatography on silica gel as described previously [4].

The amide of PGE<sub>1</sub> and the ethanolamine plasmalogen analog of PAF of the chemical structure



was synthesized by condensation of compound 1 (44  $\mu\text{moles}$ ) with PGE<sub>1</sub> (8.5  $\mu\text{moles}$ ) and N,N-dicyclohexylcarbodiimide (29  $\mu\text{moles}$ ) in the presence of chloroform (2 ml) and argon at room temperature for 20 h. After termination of the reaction, the mixture was purified by TLC using the solvent system chloroform-methanol (9:1 v/v).

The physicochemical characteristics of PGE<sub>1</sub>-PPAF are:  $R_f = 0.13$  in  $\text{CHCl}_3\text{O}-\text{CH}_3\text{OH}-\text{AcOH}-\text{H}_2\text{O}$

(130:12:4:2 v/v); IR-spectrum ( $\nu_{\text{max}}$ ,  $\text{cm}^{-1}$ ): 3450, 3050, 2825, 2860, 1740, 1650, 1550, 1470, 1240, 1065; UV-spectrum ( $\lambda_{\text{max}}$ , nm ( $\epsilon$ ) in  $\text{CH}_3\text{OH}$ ): 275.6 ( $3.8 \cdot 10^4$ ). The ratio PGE<sub>1</sub>/P<sub>i</sub> (mole per g-atom) was  $1.1 \pm 0.1$ . The PGE<sub>1</sub> content in the PGE<sub>1</sub>-PPAF was determined as described previously [10]. Lipid phosphorus was analyzed as described by Vaskovsky et al. [11].

Platelets were isolated from blood of healthy donors with 3.8% trisodium citrate solution as anticoagulant (blood/anticoagulant ratio 9:1). Platelet-rich plasma was obtained by centrifugation of blood samples at 200g for 15 min. PAF-, ADP-, or thrombin-induced platelet aggregation was measured turbidimetrically in platelet-rich plasma at 520 nm as described previously [12].

Results given in Tables 1 and 2 are the means of three parallel measurements  $\pm$  the standard error of the means. Similar results were obtained in 4-5 independent experiments.

## RESULTS AND DISCUSSION

The results of the study on the influence of PGE<sub>1</sub>-PPAF on PAF-, ADP-, or thrombin-induced platelet aggregation are presented in Table 1. Pretreatment of platelets by PGE<sub>1</sub>-PPAF ( $10^{-6}$  M) results in a complete inhibition of PAF-, ADP-, or thrombin-induced platelet aggregation. To determine whether these effects depend on the PGE<sub>1</sub> residue, we compare the influence of compound 2 having a palmitic acid residue instead of PGE<sub>1</sub> and compound 1, an ethanolamine plasmalogen PAF analog having a free amino group, on platelet aggregation. The data in Table 1 show that compounds 1 and 2 inhibit PAF-induced aggregation to some extent but has no influence on ADP- or thrombin-induced aggregation. The data on the dependence of inhibition of platelet aggregation on the concentrations of PGE<sub>1</sub>-PPAF are presented in Table 2. The data show that PGE<sub>1</sub>-PPAF inhibits PAF-induced aggregation at concentrations as low as  $10^{-9}$  M, and 100%-inhibition was observed at  $10^{-7}$ - $10^{-6}$  M PGE<sub>1</sub>-PPAF. The curves of PAF-induced platelet aggregation are presented in the figure. The data in the figure show that PGE<sub>1</sub>-PPAF at concentration of  $10^{-8}$  M significantly decreases the extent of platelet aggregation; this represents a reverse aggregation. At PGE<sub>1</sub>-PPAF concentrations of  $10^{-7}$  and  $10^{-6}$  M there is complete inhibition of the platelet aggregation, and the first wave of aggregation is not observed. PGE<sub>1</sub>-PPAF has a less inhibitory effect on the ADP-induced aggregation than on the PAF-induced aggregation, and 100%-inhibition is observed at  $10^{-6}$  M PGE<sub>1</sub>-PPAF (Table 2). Inhibition of the thrombin-induced aggregation by PGE<sub>1</sub>-PPAF revealed some peculiarities (see the figure). There are no first waves of aggregation and formation of platelet aggregates in the presence of PGE<sub>1</sub>-PPAF, and

**Table 1.** Influence of PGE<sub>1</sub>-PPAF, 1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phosphoethanolamine (compound 1), and 1-O-alk-1'-enyl-2-acetyl-sn-glycero-3-phospho-(N-palmitoyl)ethanolamine (compound 2) on the human platelet aggregation

Tested compound*	PAF	ADP	Thrombin
	inhibition of aggregation (%)		
Compound 1	50 ± 5	0	0
Compound 2	20 ± 3	0	0
PGE <sub>1</sub> -PPAF	100 ± 3	100 ± 5	100 ± 5

\* The concentration of the tested compounds was 10<sup>-6</sup> M; the concentrations of the proaggregatory agents were: PAF, 10<sup>-6</sup> M; ADP, 10<sup>-5</sup> M; thrombin, 0.1 U/ml.

**Table 2.** Inhibition of PAF- and ADP-induced aggregation of human platelets by PGE<sub>1</sub>-PPAF in platelet-rich plasma

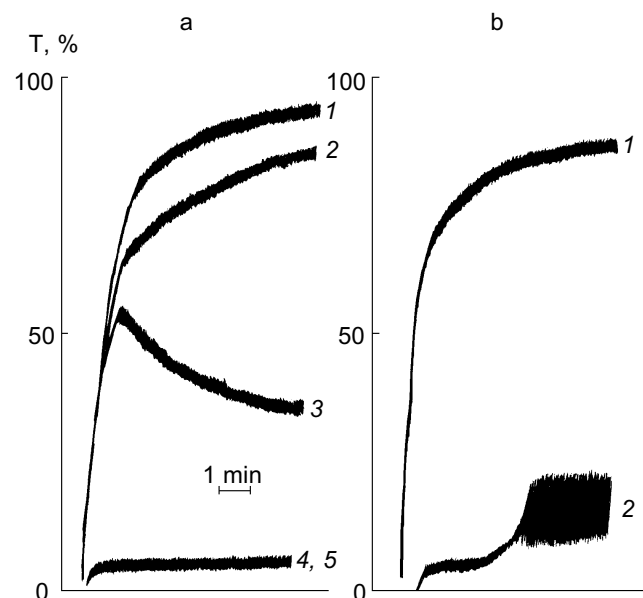
Proaggregatory agents	PGE <sub>1</sub> -PPAF concentration (M)			
	10 <sup>-9</sup>	10 <sup>-8</sup>	10 <sup>-7</sup>	10 <sup>-6</sup>
	Extent of aggregation inhibition (%)			
PAF	14 ± 3	40 ± 7	100 ± 5	100 ± 5
ADP	0 ± 2	12 ± 2	36 ± 4	100 ± 5

after a definite time (3-4 min) a sharp uneven change in light transmission is observed, indicating the formation of a jelly-like clot in the medium. The data show that PGE<sub>1</sub>-PPAF has an inhibitory effect on PAF-, ADP-, and thrombin-induced platelet aggregation. What are the reasons for these effects?

It is obvious that inhibition of PAF-induced platelet aggregation by PGE<sub>1</sub>-PPAF depends on the presence of the PGE<sub>1</sub> residue since compound 1 lacking PGE<sub>1</sub> and compound 2 having a palmitic acid residue instead of PGE<sub>1</sub> are not able to provide 100%-inhibition of platelet aggregation. The inhibitory effect of PGE<sub>1</sub>-PPAF has clear concentration dependence. It was shown previously that PGE<sub>1</sub>-stimulated cAMP formation in platelets occurs in parallel with the increase in specific binding of PGE<sub>1</sub> to platelets. The maximal cAMP formation in platelets was observed as the level of specific PGE<sub>1</sub> binding reached 0.5-1 μM [13]. These data suggest that PGE<sub>1</sub>-PPAF can act on the platelets in at least two ways: 1) to block PAF-receptors and PAF-mediated platelet activation; 2) PGE<sub>1</sub>-PPAF binds with PGE<sub>1</sub>-receptors, resulting in adenylate cyclase activation and subsequent inhibition of platelet aggregation.

ADP as well as the other proaggregatory agents induce platelet activation that involves platelet shape changes, exposure of fibrinogen receptors, aggregation, and inhibition of PGE<sub>1</sub>- or prostacyclin-stimulated adenylate cyclase [14]. It is believed that ADP-induced platelet activation and adenylate cyclase inhibition may be relatively independent events and mediated by different ADP-receptors [14]. However, changes in platelet shape and aggregation were observed at ADP concentration of 0.1-0.5 and 1-5 μM, respectively, whereas half-maximal inhibition of adenylate cyclase was observed at ADP concentration 3 μM [14]. According to our data, complete inhibition of ADP-induced aggregation is observed at 1 μM PGE<sub>1</sub>-PPAF, closely corresponding to the concentration range of ADP where inhibition of adenylate cyclase was observed. Since the synthesis of endogenous PAF is not followed by ADP-induced platelet aggregation, it may be assumed that the inhibition of ADP-induced platelet aggregation by PGE<sub>1</sub>-PPAF depends on adenylate cyclase activation and increasing platelet cAMP level.

It is known that thrombin at very low concentrations induces a reverse platelet aggregation, and at higher concentrations (0.1-1 U/ml) thrombin induces a clas-



Influence of PGE<sub>1</sub>-PPAF on PAF- or thrombin-induced platelet aggregation. a) PAF-induced aggregation: 1) platelet aggregation induced by 10<sup>-6</sup> M PAF; 2-5) platelet aggregation after pretreatment (4-5 min) of platelets with PGE<sub>1</sub>-PPAF (concentrations of PGE<sub>1</sub>-PPAF in the medium: 10<sup>-9</sup> M (2), 10<sup>-8</sup> M (3), 10<sup>-7</sup> M (4), or 10<sup>-6</sup> M (5)). b) Thrombin-induced aggregation: 1) platelet aggregation induced by thrombin (0.1 U/ml); 2) platelet aggregation after pretreatment of platelets with PGE<sub>1</sub>-PPAF followed by addition of thrombin (0.1 U/ml). Ordinate, light transmission, T (%); abscissa, time (min).

sical two-phase aggregation followed by a reverse reaction in the course of the second phase [14]. Thrombin-induced platelet activation begins with its binding to high-affinity receptors and is followed by the inhibition of platelet adenylate cyclase [14]. In our study, we found the inhibition of the first and second phase of thrombin-induced aggregation by PGE<sub>1</sub>-PPAF and observe the formation of a jelly-like fibrin clot. The reason for the formation of a fibrin clot in the absence of aggregation is unclear, but this effect appears to be linked to stimulation of the release reaction in platelets under these conditions.

Are there some other mechanisms of inhibition of platelet aggregation by PGE<sub>1</sub>-PPAF? The formation of PGE<sub>1</sub> ethanolamide from the PGE<sub>1</sub>-PPAF in the incubation medium is an alternative mechanism. Recently the inhibition of adrenaline- or arachidonic acid-induced platelet aggregation by arachidonic acid amides with dopamine, histamine, or serotonin was described [1]. It has been shown previously that amide PGE<sub>2</sub> with phosphatidylethanolamine slowly hydrolyzed in human plasma, and for 10-min incubation no more than 2% of PGE<sub>2</sub> release was observed [5]. For this reason, under our experimental conditions the possibility of PGE<sub>1</sub>-PPAF hydrolysis with free PGE<sub>1</sub> release, where all inhibitory effect is observed by 2-5 min after addition of PGE<sub>1</sub>-PPAF to platelet-rich plasma, seems unlikely. The formation of PGE<sub>1</sub> ethanolamide as the result of the action of membrane-bound phospholipase D on PGE<sub>1</sub>-PPAF appears possible. There is a thrombin-stimulated platelet phospholipase D, and its maximal activation is observed for 10 min after the action of thrombin on platelets [15].

The activity of ethanolamides of prostaglandins with respect to adenylate cyclase is unknown, whereas the ethanolamides of polyunsaturated fatty acids inhibit adenylate cyclase of various cells [1]. Thus, the data presented above show that inhibition of platelet aggregation by PGE<sub>1</sub>-PPAF could be accomplished by several mechanisms, including its action on the PGE<sub>1</sub>- and

PAF-receptors as well as the formation of ethanolamide of PGE<sub>1</sub>. The data presented suggest the possibility of simultaneous action on the platelet receptor system (PAF-receptor)-mediated platelet activation and the receptor system (adenylate cyclase) that takes part in the inhibition of platelet activation.

## REFERENCES

1. Bezuglov, V. V., Bobrov, M. Yu., and Archakov, A. V. (1998) *Biochemistry (Moscow)*, **63**, 22-27.
2. Ueda, N., Yamamoto, K., Yamamoto, S., Tokunaga, T., Shirakawa, E., Shinkai, H., Ogawa, M., Sato, T., Kudo, I., and Inoue, K. (1995) *Biochim. Biophys. Acta*, **1254**, 127-134.
3. Natarajan, V., Reddy, P. V., Schmid, P. C., and Schmid, H. H. O. (1981) *Biochim. Biophys. Acta*, **664**, 445-448.
4. Kuznetsova, T. I., and Kulikov, V. I. (1992) *Biokhimiya*, **57**, 16-20.
5. Lekim, D., Stieck, A., Betzing, H., and Kunze, H. (1978) *Advances in Prostaglandin and Thromboxane Research*, **3**, 193-198.
6. Vargaftig, B. B., Chignard, M., and Benveniste, J. (1981) *Biochem. Pharmacol.*, **30**, 263-271.
7. Kulikov, V. I., and Muzya, G. I. (1999) *Biochemistry (Moscow)*, **64**, 631-635.
8. Harby, J. H. (1999) *J. Biol. Chem.*, **274**, 7599-7602.
9. Demopoulos, C. A., Pinckard, R. N., and Hanahan, D. J. (1979) *J. Biol. Chem.*, **254**, 9355-9358.
10. Bergelson, L. D., Dyatlovitskaya, E. V., Molotkovsky, Yu. G., Batrakov, S. G., Barsukov, L. I., and Prokazova, N. V. (1981) *Preparative Lipid Biochemistry* [in Russian], Nauka, Moscow, p. 88.
11. Vaskovsky, V. E., Kostetsky, E. Y., and Vasendin, I. M. (1975) *J. Chromatogr.*, **114**, 129-141.
12. Muzya, G. I., and Kulikov, V. I. (1996) *Biochemistry (Moscow)*, **61**, 337-340.
13. Kahn, N. N., and Sinha, A. K. (1988) *Biochim. Biophys. Acta*, **972**, 45-53.
14. Siess, W. (1989) *Physiol. Rev.*, **69**, 58-178.
15. Rubin, R. (1988) *Biochem. Biophys. Res. Commun.*, **156**, 1090-1096.